Cardiodepressive Effect of Eugenyl Acetate in Rodent Heart

Leisiane Pereira Marques,1 Samuel Santos Beserra,2 Danilo Roman-Campos,3 Antonio Nei Santana Gondim1,2
Universidade do Estado da Bahia - Departamento de Educação,1 Salvador, BA - Brazil
Universidade Federal de São Paulo – Biofísica,2 São Paulo, SP – Brazil

Introduction
Cardiovascular diseases are a public health problem being among the leading causes of death worldwide. In this perspective, there is a growing interest in the search for new substances with pharmacological actions on the cardiovascular system, including drugs of natural origin.

The Syzygium aromaticum (L.) Merr. & L.M.Perry, popularly known as clove, produces several chemical compounds that have a wide range of biological effects. Eugenol (Figure 1A) is the most abundant bioactive compound found in clove essential oil, followed by eugenyl acetate (EA) (Figure 1B).

Studies have shown that eugenol has cardiodepressor activity in rats1 and guinea pigs2 probably due to the inhibition of the L-type Ca2+ current (I_{Ca,L}). In addition, eugenol acts as cardioprotector.3

Although several studies have addressed the pharmacological properties of eugenol on mammalian heart, so far there is no information on the action of EA on the cardiomyocytes. Thus, this study describes the effects of EA on atrial contractility and its inhibitory action on I_{Ca,L}.

Methods

Animals
For contractility experiments guinea pigs (Cavia porcellus, both sexes, 400-600g) were used. For electrophysiological studies, adult male C57Bl/6J mice were used. All procedures were approved by the Ethics Committee for the Use of Animals (CEUA) of the University of the State of Bahia (license: 03/2017).

Experimental Protocols

Evaluation of EA Inotropic Effect
The left atrium from guinea pig was maintained in a modified Tyrode’s solution (10 mL, 36.5±0.5°C) with the following composition (in mM): 140 NaCl; 5.4 KCl; 0.5 MgCl2; 0.33 NaH2PO4; 5 HEPES and 1.8 CaCl2 (pH=7.4), aerated with oxygen (99.9%). The atrium was stretched to attain a resting tension of 1gF and it was electrically stimulated (2Hz, 100V, 15ms). The contractile force was captured by an isometric transducer. Signals were digitized (512Hz) and stored on a computer. The left atria were subjected to increasing concentrations of EA (1-5,000μM, 3 to 5 minutes for each tested concentration).

Dimethyl sulfoxide (DMSO) was used to make the stock solution of EA (obtained from Sigma-Aldrich).

Investigation of EA Effect on the L-type Calcium Current.
Ventricular cardiomyocytes from C57BL/6J mice were isolated as previously described.4 The composition of the internal solution (in mM) was: 120 CsCl, 10 HEPES, 5 EGTA, 20 TEA-Cl, and 5 NaCl (pH=7.2; CsOH). Modified Tyrode was used as external solution. To measure L-type Ca2+ current (I_{Ca,L}) patch-clamp technique in whole-cell voltage-clamp mode was used.5,6 Cells were maintained at a resting membrane potential of -80mV, then the sarcolemma was subjected to a pre-pulse to -40mV (50ms) followed by a pulse to 0 mV (300ms, 0.1Hz). The I_{Ca,L} amplitude was measured by the difference between the current at the end of the test pulse (0mV) and the peak current. The cells were exposed to EA (10-3,000 μM, 2-3 minutes for each concentration). The signals were digitized (5kHz) and stored on the computer.

Statistical Analysis
The results are expressed as mean ± standard error of the mean and were statistically analyzed using the two-tailed “t” test (significance level: p<0.05).

Results

EA Effect on the Left Atrial Contractility
Representative traces in Figure 1C show that EA (700 μM) reduced the amplitude of atrial contraction by approximately 60% when compared to the control. This effect was partially reversed (approximately 75%) after removing the drug (washout). In Figure 1D, it is possible to observe the EA concentration-effect curve on contractility (n = 4). EA presented an IC50 (concentration that induces half of the maximum effect) of 558±24.06μM and a maximum effect of 50%.

EA Action on I_{Ca,L}
Figure 2A depicts a representative trace of the I_{Ca,L} measured in healthy isolated cardiomyocytes. Figure 2B shows the time
The cardiomyocyte contraction correlates with the increase in cytosolic Ca\(^{2+}\) concentration which is determined by the Ca\(^{2+}\) influx through Ca\(^{2+}\) channels present in the sarcolemma, as well by the amount of Ca\(^{2+}\) released by the sarcoplasmic reticulum (SR) in the process called excitation-contraction coupling. The membrane depolarization leads to the opening of the L-type Ca\(^{2+}\) channels mainly during the plateau phase of action potential leading to an inward Ca\(^{2+}\) current. This Ca\(^{2+}\) entry stimulates the release of Ca\(^{2+}\) stored in SR, a process known as Ca\(^{2+}\)-induced Ca\(^{2+}\) release, which contributes to cardiac contractility. Thus, the mechanisms that alter the intracellular Ca\(^{2+}\) handling are involved in the control of cardiac contractility.

In order to explain the negative inotropic effect of EA on the cardiac muscle, the drug effect on the I\(_{\text{Ca,L}}\) amplitude was investigated in isolated cardiomyocytes. The findings described here show that EA reduces the peak of I\(_{\text{Ca,L}}\). This effect may be associated with the activation of receptors that modulate I\(_{\text{Ca,L}}\), and/or direct action of EA on Ca\(_{\text{V1.2}}\). The reduction of I\(_{\text{Ca,L}}\) is able to explain the decrease of contractility induced by EA, since it would lead to a reduction in the release of Ca\(^{2+}\) from SR. Although additional mechanism may also be involved.
Sensch et al., studying the pharmacological properties of EA analogue eugenol, demonstrated that this substance depresses the force of atrial contraction by reduction Ca\(^{2+}\) influx in cardiomyocytes. In these experiments, it was observed that eugenol has an IC\(_{50}\) of 127\(\mu\)M, a value lower than the IC\(_{50}\) observed for AE (1,337\(\mu\)M). These data suggest that blocking effect of eugenol on Ca\(^{2+}\) channels is more potent than EA.

**Conclusion**

Eugenyl acetate has a cardiodepressor effect that can be explained, at least in part, by the inhibition of Ca\(_{v}\)1.2.

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**Author Contributions**

Conception and design of the research and Statistical analysis: Marques LP, Gondim ANS; Acquisition of data: Marques LP, Beserra SS, Gondim ANS; Analysis and interpretation of the data: Marques LP, Roman-Campos D, Gondim ANS; Obtaining financing and Critical revision of the manuscript for intellectual content: Roman-Campos D; Writing of the manuscript: Roman-Campos D, Gondim ANS.

**Potential Conflict of Interest**

No potential conflict of interest relevant to this article was reported.

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**Study Association**

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References


